

THE INFLUENCE OF N^ω-NITRO-L-ARGININE METHYL ESTER ON ARGININE AND POLYAMINE METABOLISM IN RAT'S BRAIN TISSUE DURING EXPOSITION TO MICROWAVE RADIATION

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Exposition to microwave radiation (MW) from mobile phones, satellite communications, radio relays, radars and microwave devices in medicine induce disturbances in different organ systems. It has been shown that MW from mobile phones induce increasing of oxidative stress and apoptosis of neurons with impairment of blood brain barrier, disturbances of memory and space orientation. Citrulline and nitric oxide -NO are products of L-arginine by NO sintase-NOS. L-ornithine and polyamines are products of L-arginine by arginase. N^ω-nitro-L-arginine methyl ester (L-NAME) competitive inhibits NOS and exerts neuroprotective effects.

The aim of this investigation was to determine the arginase, PAO and DAO activity, concentration of citrulline, as well as the effects of L-NAME on arginine and polyamine metabolism in brain tissue of rats exposed to MW.

Four groups of Wistar rats were investigated during 60 days: I-control-sham exposed, II (L-NAME)-rats treated with L-NAME (5 mg/kg b.w. i.p.), III (MW)-rats exposed to MW (4 h/day), IV (MW + L-NAME). The source of MW was mobile test telephone.

Decreasing activity of arginase (0.19 ± 0.04 vs. 0.25 ± 0.05 mmol/mg prot; $p < 0.01$) and increasing of citrulline concentration (10.34 ± 0.49 vs. 7.83 ± 0.41 mmol/mg prot; $p < 0.001$) were registered in the brain of MW exposed rats compared to controls. In L-NAME group there was a decrease of citrulline level ($p < 0.05$), and increase in arginase activity ($p < 0.05$) compared to controls. In the brain of exposed rats, the activity of PAO was significantly increased, while the activity of DAO was significantly increased vs. controls (1.12 ± 0.10 vs. 0.79 ± 0.09 U/mg prot; $p < 0.001$ and 0.51 ± 0.06 vs. 0.65 ± 0.06 U/mg prot; $p < 0.05$, prospectively). In MW+L-NAME group we registered increasing of DAO activity (0.61 ± 0.04 vs. 0.51 ± 0.06 U/mg prot; $p < 0.05$) in the brain tissue compared with MW group.

Having considered the obtained results, we concluded that L-NAME exerted neuroprotective effects by preventing polyamine and arginine metabolism disturbances in the rats' brain under exposition to MW. *Acta Medica Medianae* 2007;46(3):5-11.

Key words: L-NAME, microwave radiation, arginine, polyamine, brain

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Introduction

Radiation presents energy transmission through space by electromagnetic waves (EW) or atomic particles. Electromagnetic waves are consisted of electric and magnetic field which are independent entity on low frequency range but in high frequency range they are combined in unique electromagnetic field (EMF). Earth and biological systems EMF are in extremely low frequency range. Microwave radiation (MW) used in mobile

telephony, satellite communication, radio broadcasting, radars, microwave heat devices and medical diathermia are range of 300 MHz to 300 GHz. Nowadays, there are more than 2.5 billion users of mobile telephones (1).

Iniciation of signal transduction after MW interaction with cell surface has important role in apoptosis, promotion of tumor growth and pathophysiology of neurodegenerative disorders such as Parkinson's and Alzheimer diseases. Exposure to MW (10 and 20 W/cm²) significantly increased expression of apoptotic Bcl-2 proteins in rats' testicular cells (2). Increased oxidative stress after after MW exposure could be one of the important mechanisms of apoptosis (3).

It is showed that MW radiation induces disturbances in ornithine decarboxylase (ODC) enzymes activity and impaired polyamines metabolism registered in cell cultures after MW exposure. Multiple variation in ODC activity and polyamine

metabolism disturbances are dependent of applied MW frequency, exposure times and tissues. Activity of ODC in muscles cells decreased within 3-4 hours after exposure to MW radiation (4), but some literature data showed the opposite results (5).

Nonthermal MW effects induces serious changes on DNA molecules and proteins in the brain of experimental animals. This effects comprises appearance of micronucleated cells in bone marrow, conformational changes of proteins and increased oxidative stress in brain tissue (6). The effects of MW on molecular level induce numerous cellular disorders such as increased leaking of calcium ions in human neuroblastoma cell cultures, reduced secretion of melatonin or unbalanced secretion in dopaminergic system (7). Many literature data indicated that MW radiation induced DNA damage in brain cells and exerts prooxidative potential which can be modified by melatonin application (8). Lai and Singh (1995) published that MW radiation induced single or double strand DNA breaks in brain cells on dose dependent manner thus exerts cancerogenic potential (9).

L-arginine is essential amino acid and precursor for protein, urea and creatinine synthesis. Nitric oxide synthetase (NOS) induced translation of arginine to citrulline and nitric oxide (NO). Except vascular endothelial cells and activated macrophages, NO is produced in central nervous system, thymus, liver, lung, kidneys, testes and gastrointestinal tract. NO is important factor in numerous cell function regulation such as vascular tone, neurotransmission and immunogenesis. L-arginine is converted into ornithine by arginase. Decarboxylation of ornithine by ODC results in putrescine synthesis. On the other side, decarboxylation of arginine by arginine decarboxylase (ADC) results in agmatine generation. It is thought that this metabolic pathway is characteristic of plant metabolism, but nowadays ADC activity is proved in different mammalian tissues. Agmatine is hydrolysed by agmatinase into putrescine or oxidised by diamine oxidase (DAO) into gamma-guanidino-butylaldehyde, which is further converted into GABA (10).

Polyamines (spermine, spermidine and putrescine) are ubiquitous aliphatic bases widely presented in biological systems. The greatest concentration of polyamines is registered in tissues with active protein synthesis (thymus, liver, pancreas). It is well known that polyamines increasing stability of double DNA helix. In interaction with nucleic acids, polyamines are involved in many vital processes of cells proliferation, differentiation as well as regeneration and repair processes (11). Polyamines exerts antioxidant activity and react with reactive oxygen species. Polyamine accumulation in cell membranes indicates their role in regulation of permeability (12).

Initial and rate limited step in biosynthesis of polyamines in animals is decarboxylation of L-ornithine under ODC activity with putrescine forming which is precursor for further synthesis of spermine and spermidine. Biosynthesis of spermine and spermidine is irreversible reaction while interconversion of polyamines back into

putrescine is done under activity of two enzymes: spermidine/spermine N1-acetyl transferase (SSAT) and polyamine oxidase (PAO), with production of toxic acetamidopropanal and H₂O₂. These two enzymes also catalyzed conversion of spermine into putrescine by analog reactions. Putrescine could undergo oxidative desamination by the enzyme diamine oxidase (DAO), with production of gamma-aminobutyraldehyde and GABA.

N ω -nitro-L-arginine methyl ester (L-NAME) is nonselective competitive inhibitor of nitric oxide synthetase (NOS), with primary effect on constitutive forms of NOS (nNOS-expressed in brain tissue and eNOS-expressed in endothelial cells) (13). In vivo study showed that application of L-NAME induced partial but permanent inhibition of nNOS. It is also proved that application of L-NAME showed neuroprotective effects and inhibit neuronal damage (14).

The aim

The aim of this investigation was to determine the arginase, PAO and DAO activity, concentration of citrulline, as well as the effects of L-NAME on arginine and polyamine metabolism in brain tissue of rats exposed to MW.

Material and methods

Experiments were performed on 28 adult male Wistar Albino rats (8–10 weeks old, 150–200 mg), bred at the Vivarium of the Institute of Biomedical Research, Medical faculty, Nis, under conventional laboratory conditions. All animals were housed collectively in polycarbonate cages 30x40x40 cm (WxLxH) and given ad libitum access to standard laboratory food and water. The housing room was maintained at 24°C with 42±5% relative humidity and had a 12–12-h light–dark cycle (light on 06:00–18:00 h). Four groups of experimental animals (each consisted of 7 animals) are included in study.

I group (control)-sham exposed with daily intraperitoneal (i.p.) application of 1.0 ml (0.9% NaCl)

II group (L-NAME)-animals with daily application (i.p.) of 5 mg/kg BW L-NAME during 60 days

III group (MW)-animals exposed to MW radiation (4h per day) during 60 days. All animals got 1.0 ml (0.9% NaCl) (i.p.) every day 30 min before exposure.

IV group (MW + L-NAME)- rats with daily i.p. application of L-NAME (5 mg/kg BW), 30 minutes before MW exposure from mobile phones (4 h per day) during 60 days.

Experimental model of MW exposure

The microwave radiation was produced by a mobile test phone (model NOKIA 3110; Nokia Mobile Phones Ltd.) connected to a Communication Test Set PCDK with PC and appropriate software module. During microwave exposure seven freely moving rats were kept in a pure (i.e. lacking any metallic fittings) plastic

cage. In the present study, an electromagnetic near-field signal for GSM (Global System for Mobile communication) at 900 MHz (magnetic field $B=4.68 \mu\text{T}$ to $8.69 \mu\text{T}$) with continuous wave, mobile phone system was used. Source of MW was situated in the plastix box in the center of cage (maximal distance from the floor was 3 cm and from the corners 28.2 cm). Electromagnetic fields parameters in cage were measured several times during experimental exposition. The whole-body specific energy absorption (SAR) rate was estimated as 0.025-0.05 W/kg using data for a rotating ellipsoidal model of a rats.

Seven animals from each group was sacrificed after 60 days. Rats were anesthetized with ketamine HCl (2 ml/kg), administered intraperitoneally (i.p.), before sacrificing and after 15h fasting period.

Preparation for biochemical analysis

For biochemical analysis the brain tissue was cut in small pieces, washed in ice-cold isotonic NaCl solution and frozen on -20°C . Brain tissue was homogenized (20% homogenisate) in ice-cold destilated water.

Biochemical analysis

The activity of arginase in brain homogenate was estimated by Porembaska and Kedra (1975) method. Enzyme activity are measured by released free ornithine concentration. This Chinard reaction is based on fact that ornithine in addition of concentrated acet acid react with ninhydrine and create coloured compound with maximum absorbance at 515 nm. Arginase activity was expressed as $\mu\text{mol}/\text{mg}$ tissue protein.

Citruline concentration, was determined spectrophotometrically using diacetyl monoxime reaction by Boyde (1980) method. Citruline concentration was expressed in $\mu\text{mol}/\text{mg}$ of protein.

The activity of PAO and DAO were determined spectrophotometrically by Bachrach and Reches (1966) method which is based on measurements of created aminoaldehyde. Spermine is used as a substrate for PAO and putrescine for DAO. Activity of PAO and DAO were expressed as U/mg tissue protein.

Determination of proteins. Brain proteins were determined according to Lowry's method (1951), using bovine serum albumin as standard.

Statistical analysis

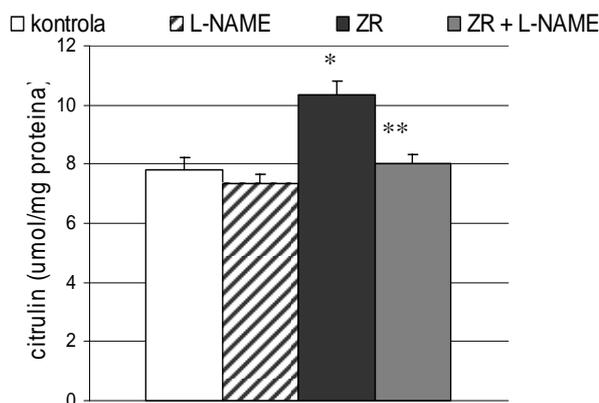
Data were analysed using a commercially available statistics software package (SPSS® for Windows, v. 9.0, Chicago, USA). Results were presented as means \pm SD. Statistical significance was determined at level of $p<0.05$ using the Student's t-test.

Results

In the MWs group, 60 days of exposure to mobile phone produces a significant decrease in

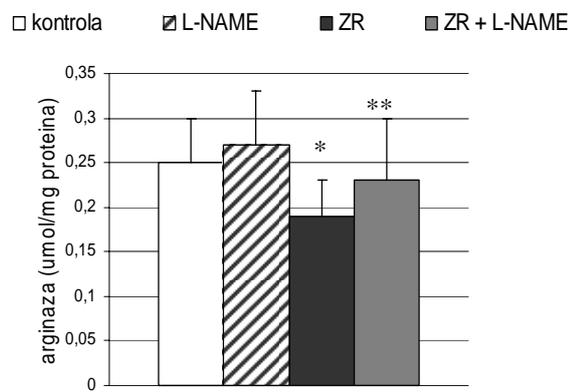
the brain tissue arginase activity ($0,19\pm 0,04$ vs. $0,25\pm 0,05 \mu\text{mol}/\text{mg}$ proteina; $p<0,01$) and increase of citruline concentration ($10,34\pm 0,49$ vs. $7,83\pm 0,41 \mu\text{mol}/\text{mg}$ proteina; $p<0,001$) when compared with control group.

Application of L-NAME to MW exposed animals (MWs+L-NAME group) significantly decreased citruline levels ($p<0,05$) and increased arginase activity ($p<0,05$) when compared with MWs groups (Graphic 1 and 2).



* $p<0.001$ (vs control);
** $p<0.05$ (vs radiation)

Graphic 1. Citruline concentration in brain of MW exposed animals

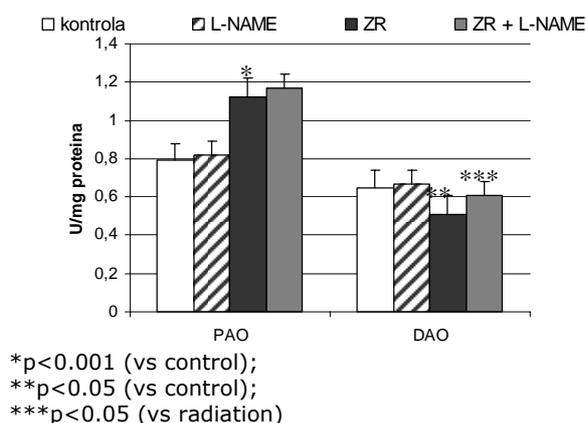


* $p<0.01$ (vs control);
** $p<0.05$ (vs radiation)

Graphic 2. Arginase activity in brain of MW exposed animals

In the brain of MW's exposed animals PAO activity is significantly higher compared to control animals ($1,12\pm 0,10$ vs. $0,79\pm 0,09$ U/mg proteins; $p<0,001$). Activity of DAO in the same tissue showed the opposite trend, and in MW's exposed animals DAO activity was significantly lower compared to control ($0,51\pm 0,06$ vs. $0,65\pm 0,06$ U/mg proteins; $p<0,05$) (Graphic 3).

Intraperitoneal application of L-NAME to animals exposed to MW (MW + L-NAME) induced moderate raise of PAO activity ($1,17\pm 0,07$ vs. $1,12\pm 0,10$ U/mg proteins; $p=ns.$) and significant raise of DAO activity ($0,61\pm 0,04$ vs. $0,51\pm 0,06$ U/mg proteins; $p<0,05$), compared to MW group (Graphic 3).



Graphic 3. PAO and DAO activity in rats brain's tissue

Discussion

The dangers from electromagnetic field (EMF) exposure by using cellular phones are huge because their effects on human health and environment are not visible in short-time period. The growing concern about possible effects on the brain after microwave (MW) exposure from mobile phones, especially in children, have been discussed in many countries. One of the first observations about negative impact of MW radiation on human health indicated many nonspecific symptoms such as irritability, neurovegetative dystonia and insomnia in workers on radar installation. It is showed that experimental animals exposed to MW expressed disorders of long term memory and orientation ability (15).

In normal conditions nitric oxide synthetase (NOS) and arginase are competitive agonists for L-arginine, converted them into citrulline+NO or polyamines. It is confirmed in the brain tissue of MW exposed rats where significant increasing of citrulline concentration was followed by decreasing of arginase activity (Graphic 1 and 2). It is most likely that MW rerout L-arginine catabolism to citrulline and NO synthesis, but not to ornithine (polyamines) synthesis.

Ilhan et al. (2004) showed that exposure to mobile telephone MW during 7 days (1 h/per day) induced significant increasing of NO (16). Significant increasing of NO in rats' retinal and brain tissues were showed after 60 days of MW exposure (17). Ozguner et al. (2005) experimentally proved significant increasing of NO in rats' myocardial cells after 10 days of mobile phones MW exposure (30 min/per day) (18).

Nitric oxide is important modulator of many physiological processes in CNS such as interneuronal communication, synaptic plasticity, releasing of neurotransmitters etc. (19). NO exerts many toxic properties by forming nitrox radicals, peroxynitrite or through NMDA system. Increased neuronal synthesis of NO contribute to cell death while increased endothelial NO synthesis exerts neuroprotective role. NOS inhibitors which antagonise eNOS and nNOS are not effective because they compromised cerebral circulation (14).

Bauer et al. (2001) showed that NO directly inhibits enzyme ornithine decarboxylase (ODC) by S-nitrosilation of sulfhydryl (SH) group in cysteine (20). It could be assumed that increased NO and citrulline production in brain of MW exposed rat, could lead to polyamine synthesis inhibition (directly by inhibition of ODC).

There are two isoforms of arginase enzyme named arginase I and arginase II. Activity of cytosolic arginase I in periportal hepatocytes contributing to urea forming, while activity of arginase II was detected in brain, kidneys and intestine (21). It is hypothesized that arginase could regulate availability of arginine for polyamine synthesis, because arginase activity is coincided with ODC and cells with arginase deficiency could not proliferate in culture without ornithine and polyamine supplementation. The great quantities of polyamines are presented in neuronal tissue (21).

It is showed that brain's cells are very sensitive on decreased polyamines concentration which is followed by compensative mechanism activation aimed to preserve normal levels of this compounds (12). In this study we registered decreasing of arginase activity and increasing of citrulline concentration, which most likely indicate that decreased synthesis of brain's polyamines was occurred under MW exposure.

There are many controversy about polyamines roles in brain's tissue. They have paradoxical role in apoptosis induction or its prevention (22). Neuroprotective roles of polyamines proceed from their anti-apoptotic and anti-oxidative properties with influencing neuronal excitability and chromatin stabilisation (23). Spermin is strong inhibitor of increased mitochondrial membrane permeability thus stopping cytochrome c leaking from mitochondria, and inhibiting apoptosis (24).

The mechanisms of neuronal damage dependent of polyamines are: 1) influx of Ca^{2+} , and neurotransmitters release in regions with increased putrescine production, 2) stimulation of NMDA receptors pathway, 3) releasing of cytochrome c from mitochondria and activation of caspase-3 with induction of apoptosis by spermine (25).

Cerebellar ODC/polyamine system is very vulnerable in pathological conditions (including brain disorders after MW exposure). It is showed that changes in polyamine metabolism connected with CNS damage have important role in neuronal degenerations (26).

Significant increasing of PAO activity in rats' brain after MW exposure (Graphic 3) could be explained by intensified polyamines interconversion, because this enzyme catalysed conversion of spermine into spermidine and spermidine into putrescine. This pathway enable regulation of polyamines levels and their disposition. During catalytic conversion spermine/spermidine into putrescine, under enzyme system SSAT/PAO, releasing of 3-acetaminopropanal and H_2O_2 is occurred. Hydrogen peroxide and aminoaldehydes, produced during polyamines degradation have cytotoxic properties. Cytotoxicity of 3-aminopropanal is regulated by acrolein forming. Toxic H_2O_2 could damage proteins, DNA and

lipida and induced apoptosis. Many literature data indicated that polyamine toxicity are the result of H₂O₂ releasing during catabolisms (27). Some polyamines metabolic intermediers induced MDA forming (8,12).

Significant oxidative damage of DNA molecules and induction of apoptosis are registred during MW exposure probably caused by increased activity of spermine oxydase (SMO) and polyamine oksidase (PAO). Increased activity of these enzymes in brain cell nucleus and citoplasm induced increasing production of H₂O₂ and OH⁻, with increased level of oxydative stress (28).

Agmatine is modulator of intracellular polyamines leves. It reduces spermine and spermidine concentration inducing activity of spermidine/spermine N¹-acetil transferase (SSAT). It is showed that agmatine induces apoptosis, by reducing cell's polyamines levels. Cultured hepatocytes supplied with agmatine showed increased number of apoptotic cells, due to increased agmatine degradation by DAO and releasing of toxic H₂O₂ (29). Galea i saradnici (1996) su dokazali da je agmatin kompetitivni inhibitor NOS, naročito njegove inducibilne izoforme (30).

Many pathological conditions are associated with changing in DAO activity. Brain's tumours are associated with decreasing activity of DAO. Rats' brain showed significant increasing in DAO activity after 60 days MW exposure compared to control animals (Graphic 3). Decreased DAO activity in MW exposed brain's tissue probably decreasing putrescine catabolism with consequent increasing of putrescine concentration. Increasing of putrescine level in brain tissue exerts neuroprotective role.

The results showed in Graohic 1 and 2 indicated that application of L-NAME decreased

citruline concentration in MW exposed rat (MW + L-NAME group) as well as increased arginase activity compared to MW exposed rats. ozračenim Having in the mind that L-NAME inhibits synthesis of NO and citruline from arginine, it is the most expected that appliacion of L-NAME would increased arginase activity. On that way, application of L-NAME, would redirected catabolism of L-arginine toward poliamine synthesis but not toward citruline and NO production. We could hypothesis that appliacion of L-NAME prevent decreasing of polyamine levels in MW exposed brain.

Increased DAO activity was registred MW exposed in rats with i.p. applicaton of L-NAME, compared to MW exposed rats (Graphic 3). It could indicate on intensified putrescine catabolic proceses in mentioned conditions (31). Having in the mind that inhibitors of NO synthesis reduced GABA releasing in the brain cortex (32), it is posible that reduced NO levels after MW exposure, activated alternative pathway of GABA synthesis from putrescine catalised by DAO, which present logical explanantion for increased activity of this enzymes.

Conclusion

Obtained results indicate that in rats brain tissue occured significant increasing of citruline and PAO activity folowed by decreasing of arginase and DAO activity. after 60 days of MW exposure. Application of L-NAME to MW exposed rats prevent disturbances in arginine metabolism by increased arginase and DAO activity, citruline and polyamines levels.

Literatura

1. Krstić D, Đinđić B, Kocić G, Petković D, Radić S, Sokolović D. Štetna delovanja elektromagnetnog polja učestanosti 50 Hz na biološke sisteme. *Acta Medica Medianae* 2003; 42(4):7-14.
2. Yu CH, Guo C, Yao YQ. Effects of high power microwave on the expressions of Bcl-2 and C-myc proteins in the rat testis. *Zhonghua Nan Ke Xue* 2005; 11(1): 22-5.
3. Djindjić B, Sokolović D, Radić S, Pavlović T, Cvetković M, Radisavljević J. Biološki efekti mikrotalasnog zračenja na moždano tkivo kod pacova. *Acta Medica Medianae* 2003; 42(2):9-12.
4. Cain CD, Thomas DL, Ghaffari M, Adey WR. 837 MHz Digital Cellular Telephone RF Fields and Induced ODC Activity in C3H10T1/2 Cells, The Bioelectromagnetics Society Meeting, Victoria, BC, Canada, 1996.
5. Paul Raj R, Behari J, Rao AR. Effects of low level 2.45 GHz microwave radiation on Ca²⁺ efflux and ODC activity in chronically exposed developing rat brain. *Natural Seminar on Low-Level Electromagnetic Field Phenomena in Biological Systems*, New Delhi, Indija, 1999.
6. Demisia G, Vlastos D and Matthopoulos DP. Effect of 910-MHz electromagnetic field on rat bone marrow. *ScientificWorld Journal* 2004; 4:48-54.
7. Burch JB, Reif JS, Noonan CW, Ichinose T, Bachand AM and Koleber TL et al. Melatonin metabolite secretion among cellular telephone users. *Int J Radiat Biol* 2002; 78:1029-36.
8. Sokolovic D, Djindjic B, Nikolic J, Bjelakovic G, Pavlovic D, Kocic G, et al. Melatonin reduces oxidative stress induced by chronic exposure of microwave radiation from mobile phones in rat brain. *J Radiat Res* 2008; 49(6):579-586.
9. Lai H, Singh NP. Acute low-intensity microwave exposure increases DNA single-strand breaks in rat brain cells *Bioelectromagnetics* 1995;16(3):207-10.
10. Grillo MA and Colombatto. Metabolism and function in animal tissues of arginine, a biogenic amine formed from arginine. *Amino Acids* 2004; 26:3-8.
11. Bjelaković G, Kocić G, Pavlović D, Nikolić J, Stojanović I, Bjelaković GB, Jevtović T, Sokolović D. Effects of Folic Acid on Polyamine Concentrations and Polyamine Oxidase Activity in Regenerating Rat Liver. *Pteridines* 2003;14: 109-113.
12. Sokolović D, Bjelaković G, Zajić S, Damnjanović Z, Nikolić J, Kocić G et al. Efekti L-metionina na metabolizam poliamina u moždanom tkivu pacova sa holestazom. *Acta Medica Medianae* 2006; 45(1):21-26.
13. Boehr R, Ulrich WR, Klein T, Mirau B, Haas S, Baur I. The inhibitory potency and selectivity of arginine substrate site nitric oxide synthase inhibitors is solely determined by their affinity toward the different isoenzymes. *Mol Pharmacol* 2000; 58(5): 1026-34.
14. Zhang ZG, Reif D, MacDonald J, Tang WX, Kamp DK, Gentile RJ, Shakespeare WC, Murray RJ, Chopp M. ARL 17477, a potent and selective neuronal NOS inhibitor decreases infarct volume after transient middle cerebral artery occlusion in rats. *J Cereb Blood Flow Metab* 1996; 16: 599-604.

15. Laurence AJ, French WP, Linder AR, McKenzie RD. Biological effects of electromagnetic fields-mechanism for the effects of pulsed microwave radiation on protein conformation. *J Theor Biol* 2000;206:291-8.
16. Ilhan A, Gurel A, Armutcu F, Kamisli S, Iraz M, Akyol O, Ozen S. Ginkgo biloba prevents mobile phone-induced oxidative stress in rat brain. *Clin Chim Acta* 2004; 340:153-62.
17. Ozguner F, Bardak Y, Comlekci S. Protective effect of melatonin and caffeic acid phenethyl ester against retinal oxidative stress in long-term use of mobile phone: A comparative study. *Molecular and Cellular Biochemistry* 2006; 282: 83-88.
18. Ozguner F, Oktem F, Ayata A, Koyu A, Yilmaz H. R. A novel antioxidant agent caffeic acid phenethyl ester prevents long-term mobile phone exposure-induced renal impairment in rat. *Molecular and Cellular Biochemistry* 2005; 277:73-80.
19. Liaudet L, Soriano FG, Szabo C. Biology of nitric oxide signalling. *Critical Care Med* 2000; 28(4):N37-N52.
20. Bauer PM, Buga GM, Fukuto JM, Pegg AE, Ignarro LJ. Nitric oxide inhibits ornithine decarboxylase via S-nitrosylation of cysteine 360 in the active site of the enzyme. *J Biol Chem* 2001; 276(37):34458-64.
21. Gotoh T, Araki M, Mori M. Chromosomal localisation of arginase II gene and tissue distribution of its mRNA. *Biochem Biophys Res Commun* 1997; 233:487-91.
22. Thomas T and Thomas TJ. Polyamines in cell growth and cell death: molecular mechanisms and therapeutic applications. *Cell Mol Life Sci* 2001; 58:244-58.
23. Adibhatla RA, Hatcher JF, Sailor K, Dempsey J. Polyamines and central nervous system injury: spermine and spermidine decrease following transient focal cerebral ischemia in spontaneously hypertensive rats. *Brain Research* 2002; 1: 30-8.
24. Stefanelli C, Stanić I, Zini M, Bonavita F, Flamigni F, Zamboni L, et al. Polyamines directly induce release of cytochrome c from heart mitochondria. *Biochem J* 2002; 347:875-80.
25. Muralikrishna Rao A, Hatcher J, Dempsey R. Polyamine response to CNS injury: for better or for worse? *Recent Research Developments in Neurochemistry* 1999; 2: 517-32.
26. Henley CM, Wey K, Takashima A, Mills C, Granmayeh E, Krishnappa I et al. S-adenosylmethionine decarboxylase activity is decreased in the rat cortex after traumatic brain injury. *J Neurochem* 1997;69:259-65.
27. Paschen W. Polyamine metabolism in different pathological states of the brain. *Mol Chem Neuropath* 1992; 16: 241-71.
28. Amendola R, Bellini A, Cervelli M, Degan P, Marecchi L, Martini F, Mariottini P. Direct oxidative DNA damage, apoptosis and radio sensitivity by spermine oxidase activities in mouse neuroblastoma cells. *Biochimica et Biophysica Acta* 2005; 1755:15-24.
29. Gardini G, Cravanzola C, Testore G, Solinas SP, Colombatto S. Agmatine inhibits the proliferation of rat hepatoma cells by modulation of polyamine metabolism. Abstracts Meeting "Biogenic Amines", Albere di Tenna, Trento, 2002, p 4P.
30. Galea E, Regunathan S, Eliopoulos V, Feinstein DL, Reis DJ. Inhibition of mammalian nitric oxide synthases by agmatine, an endogenous polyamine formed by decarboxylation of arginine. *Biochem J* 1996;316: 247-49.
31. Wallace H, Fraser A, Hughes A. A perspective of polyamine metabolism. *Biochem J* 2003; 376:1-14.
32. Montague PR, Gancayco CD, Winn MJ, Marchase RB, Friedlander MJ. Role of NO production in NMDA receptor-mediated neurotransmitter release in cerebral cortex. *Science* 1994; 263: 973-7.

UTICAJ N^ω-NITRO-L-ARGININ METIL ESTRA NA METABOLIZAM ARGININA I POLIAMINA U MOŽDANOM TKIVU PACOVA U TOKU IZLAGANJA MIKROTALASNOM ZRAČENJU

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Izlaganje mikrotalasnom zračenju (MW), koje se koristi za mobilnu telefoniju, satelitsku komunikaciju, radio emitere, radare, mikrotalasne toplotne uređaje i medicinsku dijatermiju dovodi do pojave poremećaja u različitim organskim sistemima. Dokazano je da MW mobilnih telefona dovodi do povećanja nivoa oksidativnog stresa i apoptoze neurona, narušavanja integriteta krvno-moždane barijere i poremećaja dugotrajne memorije i sposobnosti orijentacije. Iz L-arginina se pod dejstvom enzima azot monoksid sintaze (NOS) stvaraju citrulin i azot monoksid (NO), a pod dejstvom arginaze nastaju L-ornitin i poliamini (neophodni za rast, proliferaciju i regeneraciju ćelija). Nω-nitro-L-arginin metil ester (L-NAME) je neselektivni kompetitivni inhibitor NOS, koji pokazuje neuroprotektivno dejstvo i sprečava oštećenje neurona.

Cilj ovog istraživanja bio je da se nakon izlaganja pacova MW mobilnog telefona, u moždanom tkivu odredi: aktivnost arginaze, količina citrulina, katabolizam poliamina (određivanjem aktivnosti PAO i DAO), kao i efekat L-NAME na metabolizam arginina i poliamina.

Wister pacovi bili su podeljeni u četiri eksperimentalne grupe: I (kontrola), II (L-NAME) – životinjama je 60 dana svakodnevno davan L-NAME (5mg/kg TM), III (ZR) – životinje su 60 dana izlagane MW (4h/dnevno), IV (ZR + L-NAME) – pacovi kojima je aplikovan L-NAME, 60 dana (4h/dnevno) su izlagani MW mobilnog telefona.

U mozgu pacova koji su bili izloženi MW zabeleženo je sniženje aktivnosti arginaze (0.19±0.04 naspram 0.25±0.05 μmol/mg proteina; p<0.01) i značajno povišenje nivoa citrulina (10.34±0.49 naspram 7.83 ±0.41 μmol/mg proteina; p<0.001), u odnosu na kontrolne životinje. Aplikovanje L-NAME zračenim pacovima dovodi do sniženja nivoa

citrulina ($p < 0.05$), kao i do povišenja aktivnosti arginaze ($p < 0.05$), u odnosu na kontrolu. Aktivnost PAO u tkivu mozga ozračenih pacova je značajno povišena, dok je aktivnost DAO značajno snižena u odnosu na kontrolne životinje (1.12 ± 0.10 naspram 0.79 ± 0.09 U/mg proteina; $p < 0.001$ i 0.51 ± 0.06 naspram 0.65 ± 0.06 U/mg proteina; $p < 0.05$). Kod pacova koji su izlagani mikrotalasnom zračenju, a kojima je aplikovana L-NAME, zapaženo je povišenje aktivnosti DAO (0.61 ± 0.04 naspram 0.51 ± 0.06 U/mg proteina; $p < 0.05$) u tkivu mozga u odnosu na ZR + L-NAME grupu.

Aplikovanjem L-NAME pacovima koji su izlagani MW dolazi do sprečavanja poremećaja metabolizma arginina i poliamina u moždanom tkivu i tako ova supstanca pokazuje neuroprotektivno dejstvo. *Acta Medica Medianae 2007;46(3): 5-11.*

Ključne reči: L-NAME, mikrotalasno zračenje, arginin, poliamini, mozak