

INFLUENCE OF VITAMIN C AND RIBWORT PLANTAIN EXTRACT ADDITION TO THE PROPOLIS EXTRACT ON THE VIABILITY OF FIBROBLASTS IN CELL CULTURE *IN VITRO*

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ABSTRACT

Propolis is a honey bee product rich in biologically active substances that are proved to have beneficial effects on human health. It is widely used in traditional medicine for the treatment of various respiratory but also skin disorders. Propolis can be used alone as pure extract or with the addition of various plant extracts and antioxidants in order to achieve synergistic effects. The aim of this study was to examine the effects of different propolis extracts, commercially available on the market, on the viability of fibroblasts in cell culture *in vitro*. We examined the effect of three different propolis extracts: pure propolis extract (25%), propolis extract (10%) with added vitamin C and propolis extract (10%) with Ribwort Plantain extract and added vitamin C, on the viability of L929 fibroblasts, using MTT test and microscopically. Concentration-dependent effect of all examined propolis extracts on the viability of fibroblasts was observed. Also, differences in the effect of examined extracts on cell viability were noticed related to the additions to the propolis extract and the pattern was different in lower compared to the higher examined concentrations. Addition of vitamin C and Ribwort Plantain extract influence the effects of pure propolis. Using propolis in combination with plant extracts and bioactive substances may have beneficial effects but it should be considered based on the indications for which these products are intended and effects to be achieved.

Key words: propolis, Ribwort Plantain, vitamin C, fibroblasts, *in vitro*

INTRODUCTION

Propolis is a natural resinous substance produced by bees from substances they collect from different parts of plants and exudates. The chemical composition of propolis, biological and pharmacological activity vary greatly depending on the plant species from which it originates and the geographical origin (1). Propolis is mainly composed of resins and vegetable balsam (50%), wax (30%), essential oils (10%), pollen (5%), sugars, amino acids, vitamins, and minerals (5%) (2-9). Except resins and waxes, the main groups of chemical compounds found in propolis are phenols (e.g., flavonoids, polyphenols, phenolic acids and other phenolic compounds) and their esters, terpenes and terpenoids, steroids, aromatic acids, aromatic esters, aldehydes, alcohols, sugars and their alcohols and acids, amino acids, vitamins, fatty acids, hydrocarbons, mineral elements and alcohols (7, 9-12). Flavonoids are a major group of phenolic compounds in propolis which greatly contribute to the biological and pharmacological activities of propolis. Propolis has been known for a long time in the traditional medicine of many countries for its therapeutic properties, so in some countries it is used to treat a wide range of diseases and is also used for cosmetic purposes (13). It is known that propolis has an antibacterial, antiviral and antifungal activity (8, 9, 13), so it is used to treat infections of the upper respiratory tract as well as superficial wounds in the form of topical formulations (14). Propolis acts as an antioxidant and anti-inflammatory agent, so it is also used in the treatment of inflammatory processes and disorders (8, 9, 13). Since propolis has been shown to have a beneficial effect on wound healing (13, 15), it is used in the form of topical formulations for healing of wounds and burns on the skin and in dental preparations for the treatment of gingivitis (16). Nowadays, propolis is used in the form of drops and sprays for oral and *per os* use, in the form of creams, gels, lotions, capsules, lozenges, toothpastes and others.

Vitamin C is a well-known antioxidant that is added to various products in order to achieve synergistic effect and to potentiate their antioxidant capacity. In addition to its antioxidant effect, vitamin C stimulates fibroblast proliferation and collagen synthesis (17), so it is often added to preparations for skin care and wound healing. In cosmetology and dermatology, vitamin C is commonly used as a component of various creams and serums because it has a protective effect on skin damage caused by various harmful effects and slows down the aging process (18). The addition of vitamin C to various formulations can contribute to and stimulate their beneficial properties.

Ribwort Plantain (*Plantago lanceolata* L.) has been known for its beneficial effects on human health since ancient time. *Plantago lanceolata* L. is a species from the genus *Plantago*, family Plantaginaceae, that is the most commonly used for medicinal purposes. Ribwort Plantain is registered as a natural medicine (herbal medicinal product) by the European Medicines Agency (EMA). EMA defines the oral, oromucosal and cutaneous use of the leaf of Ribwort Plantain (lat. *Plantaginis lanceolatae folium*) for the following indications: as a demulcent for the symptomatic treatment of oral or pharyngeal irritations and associated dry cough, for the relief of cough associated with cold and for the treatment of minor inflammation of the skin (19).. In many countries, it is used in combination with other plant species to treat various disorders. It is used in the form of extracts, syrups, teas, lozenges and tablets by oral and topical route for the treatment of inflammation of the upper and lower parts of the respiratory tract, inflammation of the skin and for the healing of skin and mucous membranes wounds (20). Ribwort Plantain has been shown to possess anti-inflammatory (21), antioxidant (22), antibacterial (23), and antiviral activity (24), to stimulate epithelization and to act as spasmolytic agent (25). It is one of the main plants used in cough remedies and exert anti-inflammatory effects, protects the liver, and is used in the treatment of cancer (26). The main biologically active compounds of *Plantago lanceolata* L. are: iridoid glycosides (aucubin and catalpol), phenylethanoid glycosides, polysaccharides, flavonoids (apigenin and

luteolin), polyphenols, alkaloids, terpenoids, fatty acids, phytosterols, phenylethanoids (acteoside, plantamajoside) and tannins, organic acids, mucilaginous substances, mineral salts, and pectins (26-29). The Ribwort Plantain extract can successfully be used as an effective ingredient in cosmetic products (30).

In biomedical research where biological activities of various substances and pharmaceutical products are examined, studies on cell cultures *in vitro* are of fundamental importance and represent the first step in examining the biological activities. In order to investigate the potential effects such as wound healing on the skin and mucous membranes, the most commonly used model for this type of investigation are fibroblast cultures. The cell line that is the most suitable for this purpose is L929 cell line, a permanent cell line of fibroblasts obtained from the mouse skin.

The aim of this study was to examine the influence of addition of vitamin C and Ribwort Plantain extract to the propolis extract on the viability of fibroblasts in cell culture *in vitro*.

MATERIALS AND METHODS

Propolis extracts

Three different propolis ethanol extracts available on the market were tested: highly purified propolis extract (25%) in ethanol solution (Propolis drops – extra 25%), highly purified propolis extract (10%) in ethanol solution with added vitamin C (Propolis drops with vitamin C), highly purified propolis extract (10%) and *Plantaginis lanceolatae folium* extract in an ethanolic solution with added vitamin C (Propolis drops with Ribwort Plantain and vitamin C), all from producer Sinefarm d.o.o. Serbia. Extracts were diluted in complete DMEM and final dilutions 1/100, 1/200, 1/500, 1/1,000 and 1/2,000 (v/v) were examined.

Cell culture and viability assay

The effect on cell viability was examined on L929 cell line (obtained from American Type Culture Collection – ATCC) which were cultured in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum, antibiotic-antimycotic solution and stable glutamine (which makes complete DMEM medium), all purchased from Capricorn Scientific GmbH, Germany. Cells were seeded in 96-well culture plates (Greiner Bio-One, Germany) in complete DMEM so that 20,000 cells were seeded per well per 100 μ l of medium, and left for 24 hours in an incubator with 5% CO₂, in humidified atmosphere, at 37°C, to adhere and to adapt to the environment. After that, 100 μ l of prepared dilutions of examined propolis extracts (twice as high as those finally tested) were added to the cells. Cells were incubated with different concentrations of examined propolis extracts for the next 24 hours and after that MTT test was performed to assess the effect of extracts on cell viability. MTT test is commonly used for viability testing and is based on the ability of viable cells to reduce the yellow tetrazolium salt MTT by mitochondrial dehydrogenases, to violet formazan crystals which are further dissolved in 2-propanol. The intensity of the colored solutions obtained from the dissolved formazan crystals is in a direct correlation with the number of viable cells. Absorbance was measured on a multichannel spectrophotometer Multiscan Ascent (ThermoLab Systems, Finland) at the wavelength 540 nm with the correction at 650 nm. Each concentration was tested in tetraplicates and experiment was repeated twice. As a control we used cells that were incubated under the same conditions in complete medium but without examined extracts (untreated cells).

Microscopical analysis

Cells were analyzed microscopically using inverted light microscope (Observer Z1, Carl Zeiss, Germany), under phase contrast. The images of cells were acquired before incubation with extracts and 24 hours after incubation, prior to MTT test, using the camera AxioCam HR in a software ZEN 2 blue edition (Carl Zeiss, Germany).

Statistical analysis of the results

The results of the MTT test are shown as a percentage of cell viability in relation to the control (untreated cells), for which the viability was considered to be 100%, with relative standard deviations. The percentage of cell viability is obtained from the mean values of the measured absorbances for each examined concentration and control. Statistically significant differences were analyzed by one-way ANOVA (analysis of variance) test and values for which $p < 0.05$ were considered as significant.

RESULTS

Effects of examined propolis extracts on the viability of L929 fibroblasts were assessed by MTT test and microscopically. Results of MTT test are shown in Figure 1.

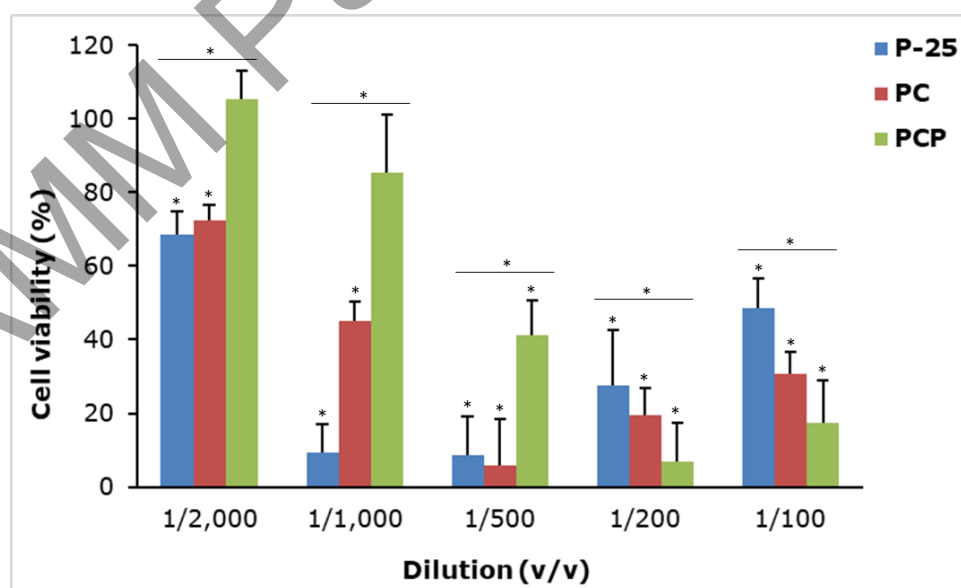


Figure 1. The effect of different propolis extracts: pure propolis extract 25% (P-25), propolis extract with added vitamin C (PC) and propolis extracts with Ribwort Plantain and added

vitamin C (PCP), on the viability of L929 cells expressed as a percentage of cell viability compared to the control (untreated cells); (*) $p < 0.001$

Statistically significant difference in the effect of examined propolis extracts on L929 cell viability was observed at all examined dilutions. Significant decrease in cell viability, i.e., cytotoxicity was noticed for all examined dilutions of P-25 and PC extracts with more pronounced cytotoxic effect of dilutions from 1/100 to 1/1,000. PCP extract was shown to be the least cytotoxic among examined extracts. Dilutions 1/1,000 and 1/2,000 of PCP extract were not cytotoxic for L929 cells. The trend of action of examined extracts in dilutions 1/100 and 1/200 is the same, the PCP extract is the most cytotoxic, while the P-25 extract is the least toxic at these concentrations. With a decrease in the concentration of all three extracts, i.e., increase in dilution, the trend of action changes, so the most cytotoxic effect is noticed for the P-25 extract, while the least cytotoxic is the PCP extract.

The effect of examined extracts on cell morphology and number was examined microscopically and the results are shown in Figure 2. In higher examined concentrations (dilutions from 1/500 to 1/100) cells were round in shape which indicate toxic effects, with some cells becoming elongated in the case of 1/500 dilution of PCP extract. At dilution 1/1,000 differences among examined extracts were noticed, with the most pronounced cytotoxic effect reflected on the cell morphology in the case of P-25 extract, followed by PC extract where there are some cells elongated in shape and PCP extract with most of the cells elongated in shape which is typical for fibroblasts. These observations are in accordance with the results of MTT test. At dilution 1/2,000 differences in cell density besides differences in cell morphology can be noticed which are in accordance with the results of MTT test in terms that PCP extract did not exert toxic effects, which is visible as the highest cell density on microscopical images and morphology of cells typical for fibroblasts.

Solvent (ethanol at dilution 1/100) did not influence the cell morphology and number of cells, nor acting cytotoxic (% of cell viability measured by MTT test was 112% which was slightly, but significantly stimulated parameter of cell viability compared to the control). This means that solvent did not cause cytotoxic effects on cells.

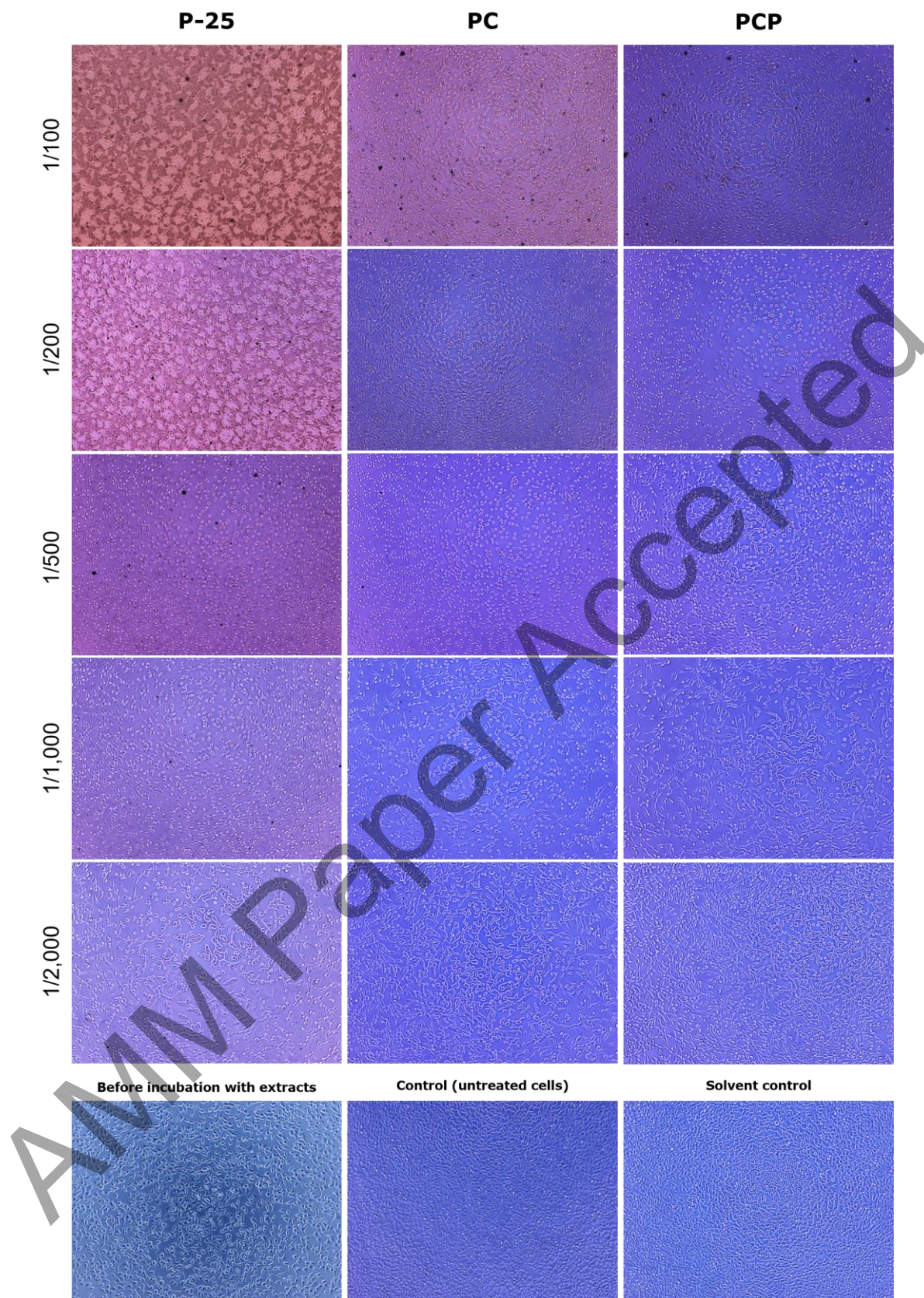


Figure 2. Morphological appearance of L929 fibroblasts before incubation with extracts, 24 hours after incubation with different dilutions of pure propolis extract 25% (P-25), propolis extract with added vitamin C (PC), propolis extract with Ribwort Plantain and added vitamin C (PCP), control (untreated cells) and solvent control.

DISSCUSSION

The obtained results show that all examined propolis extracts exhibit a concentration-dependent effect. A different effect on cell viability was obtained in the treatment of cells with different propolis extracts. At higher dilutions of the extracts, e.g., lower concentrations of the extracts, the protective effect of vitamin C and Ribwort Plantain extract on cell viability was pronounced, compared to the effect of pure propolis extract. At lower dilutions, i.e., higher concentrations of extracts, the cytotoxic effect is present regardless of the type of extract used. In order to rule out the potential effect of the solvent used for extract preparation, the cells were incubated with the same dilutions of ethanol, which was used as a solvent in all examined extracts, where even the highest concentration of ethanol, a dilution of 1/100, was not cytotoxic for L929 fibroblasts, moreover, it slightly increased the reduction of MTT by fibroblasts. Concentration of ethanol in all examined dilutions of propolis extracts was less than 1%.

The cytotoxic and antiproliferative effect of propolis and extracts of *Plantago* spp. was demonstrated on cancer cell lines, which gives these extracts a potential application in chemotherapy (31, 32). This effect was mainly demonstrated on epithelial cancer cells.

The ethanolic extracts of Malaysian and Brazilian red propolis show potential to assist in wound healing, depending on their concentration, in an *in vitro* cell model using normal human fibroblast cell line CRL-7522 (33). In the study where Portuguese 30% propolis ethanolic extract was examined on human dermal fibroblasts and keratinocytes, it was found that concentrations of propolis extract below 1 mg/mL was well-tolerated by fibroblasts and moderately tolerated by keratinocytes, which, together with good antimicrobial and antibiofilm effect, suggests that propolis extract could have a good applicability in the form of topical formulations for antibacterial treatment of infected skin disorders (14).

Vitamin C is important for collagen synthesis by fibroblasts and is very popular addition to the creams and other topical formulations to prevent skin damage and aging but also in different products to prevent gum retraction. In a study where human gingival fibroblasts were rinsed with 0, 10, 20 and 50 µg/mL of L-ascorbic acid for 7 min, three times per day in the experiment that lasted for two days, it was shown that rinsing the fibroblasts with 50 µg/mL of L-ascorbic acid significantly reduced the cell viability, evaluated by MTT test, while concentrations below 50 µg/mL did not influence the cell viability (34). In our study, propolis extract with added vitamin C (PC) led to decreased cell viability at dilutions from 1/100 to 1/500, compared to the pure propolis extract (P-25) without vitamin C. The concentration of vitamin C in extracts PC and PCP examined in our study, was 3 g/100 mL according to the data available on the website of the manufacturer, which means that 1/500 dilution of PC extract contains 60 µg/mL of vitamin C which is close to the concentration of vitamin C used in the mentioned study. In lower concentrations (dilutions 1/1,000 and 1/2,000) propolis extract with added vitamin C was shown to be less cytotoxic compared to the pure propolis extract. The important difference was also that in the mentioned study fibroblasts were only rinsed with vitamin C, while in our study fibroblasts were incubated for 24 hours with propolis extracts containing vitamin C. Also, we used permanent cell line, while in the mentioned study primary culture of gingival fibroblasts was used.

Extracts from *Plantago ovata* has been shown to have a beneficial effect and to stimulate the proliferation of fibroblasts *in vitro* (35). Extracts of *Plantago lanceolata* L. were shown to be a valuable source of bioactive substances that have beneficial effects on fibroblasts due to high antioxidant properties, UV protecting activity and stimulation of skin regeneration, which makes those extracts a good additive to the pharmaceutical formulations that are used as natural cosmetics (26). Wound healing activity of aqueous extract of Ribwort Plantain was

demonstrated in an open wounds rat model (36). It was also shown that different solvents influence different effects of the Ribwort Plantain extracts on fibroblasts. For example, extracts using glycol and glycerin allowed isolation of some bioactive compounds that were not present in ethanolic and aqueous extracts while ethanolic extract of Ribwort Plantain at concentrations of 50 mg/mL reduced the viability of fibroblasts by 45% (26). In our study, propolis ethanolic extract with Ribwort Plantain and added vitamin C was examined and was shown to be the least cytotoxic among examined propolis extracts, with absent of cytotoxic activity at lower examined concentrations (1/1,000 and 1/2,000), while pure propolis extract and propolis extract with added vitamin C were cytotoxic at those dilutions.

There are numerous propolis extracts on the market. Differences in the action of propolis extracts that can be found in the literature can be explained by different composition of propolis from different countries, which is inevitable, because the composition of propolis depends primarily on the type of plants and their parts from which bees collect it. Combining propolis extract with plant extracts may have beneficial effects on human health if combination is properly designed. It is also very important to consider the potential application of propolis extracts prior to mixing with other biologically active compounds. Further studies with propolis extracts that we examined are necessary, in order to analyze the effects of those extracts on wound healing both *in vitro* and *in vivo*, before making any conclusions regarding the potential use of those extracts in the treatment of skin disorders.

CONCLUSIONS

Based on the obtained results, it can be concluded that there is a concentration-dependent effect of all examined propolis extracts and that there is a difference in the effect of the extracts in relation to the composition. Addition of vitamin C to the propolis extract as well as combination of propolis extract with Ribwort Plantain change the effect of propolis on the viability of fibroblasts in cell culture. When choosing the propolis extracts with additions of other extracts or vitamin C, potential indication and application of that extract should be considered. Further *in vitro* and *in vivo* studies are needed before recommendations on the use of these combinations in the treatment of skin disorders.

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REFERENCES

1. Ghisalberti EL. Propolis: a review. *Bee World* 1979; 60:59–84.
2. Burdock GA. Review of the biological properties and toxicity of bee propolis (propolis). *Food and Chemical Toxicology* 1998; 36(4):347–363.
3. Park YK, Alencar SM, Aguiar CL. Botanical origin and chemical composition of Brazilian propolis. *Journal of Agricultural and Food Chemistry* 2002; 50(9):2502–2506.
4. Pietta PG, Gardana C, Pietta AM. Analytical methods for quality control of propolis. *Fitoterapia* 2002; 73(1):S7–S20.

5. Ristivojević P, Trifković J, Andrić F, Milojković-Opsenica D. Poplar-type Propolis: Chemical Composition, Botanical Origin and Biological Activity. *Nat Prod Commun* 2015; 10(11):1869-76.
6. Gómez-Caravaca A, Gómez-Romero M, Arráez-Román D, Segura-Carretero A, Fernández-Gutiérrez A. Advances in the analysis of phenolic compounds in products derived from bees. *J Pharmaceut Biomed* 2006; 41(4):1220–1234.
7. Huang S, Zhang C-P, Wang K, Li GQ, Hu F-L. Recent advances in the chemical composition of propolis. *Molecules* 2014; 19(12):19610–19632.
8. Sforcin JM. Biological Properties and Therapeutic Applications of Propolis. *Phytother Res* 2016; 30(6):894-905.
9. Pasupuleti VR, Sammugam L, Ramesh N, Gan SH. Honey, Propolis, and Royal Jelly: A Comprehensive Review of Their Biological Actions and Health Benefits. *Oxid Med Cell Longev* 2017; 2017:1259510.
10. Bankova V, Popova M, Trusheva B. Propolis volatile compounds: chemical diversity and biological activity: a review. *Chem Cent J* 2014; 8:28.
11. Madrigal-Santillán E, Madrigal-Bujaidar E, Álvarez-González I, Sumaya-Martínez MT, Gutiérrez-Salinas J, Bautista M, et al. Review of natural products with hepatoprotective effects. *World J Gastroenterol* 2014; 20(40):14787-14804.
12. Walker P, Crane E. Constituents of propolis. *Apidologie* 1987; 18:327–334.
13. Stojanović S, Najman SJ, Bogdanova Popov B, Najman SS. PROPOLIS: CHEMICAL COMPOSITION, BIOLOGICAL AND PHARMACOLOGICAL ACTIVITY – A REVIEW. *Acta Medica Medianae*, 2020; 59(2):108-113.
14. Queiroga MC, Laranjo M, Andrade N, Marques M, Costa AR, Antunes CM. Antimicrobial, Antibiofilm and Toxicological Assessment of Propolis. *Antibiotics*. 2023; 12(2):347.
15. Martinotti S, Ranzato E. Propolis: a new frontier for wound healing? *Burns Trauma* 2015; 3:9.
16. Wagh VD. Propolis: a wonder bees product and its pharmacological potentials. *Adv Pharmacol Sci*. 2013; 2013:308249.
17. Hata R-I, Senoo H. L-ascorbic acid 2-phosphate stimulates collagen accumulation, cell proliferation, and formation of a three-dimensional tissue-like substance by skin fibroblasts. *J Cell Physiol*. 1989; 138:8–16.
18. Farris PK. Topical Vitamin C: A Useful Agent for Treating Photoaging and Other Dermatologic Conditions. *Dermatologic Surgery* 2005; 31:814–818.
19. European Medicines Agency. *Plantaginis lanceolatae folium* - herbal medicinal product, Ribwort Plantain. European Medicines Agency, 2007. <https://www.ema.europa.eu/en/medicines/herbal/plantaginis-lanceolatae-folium> (Accessed on 2024-11-10)
20. Bahramsoltani R, Farzaei MH, Rahimi R. Medicinal plants and their natural components as future drugs for the treatment of burn wounds: an integrative review. *Arch Dermatol Res*. 2014; 306(7):601-617.
21. Herold A, Cremer L, Calugaru A, Tamas V, Ionescu F, Manea S, Szegli G. Hydroalcoholic plant extracts with anti-inflammatory activity. *Rom Arch Microbiol Immunol* 2003; 62(1-2):117-129.

22. Gálvez M, Martín-Cordero C, Houghton PJ, Ayuso MJ. Antioxidant activity of methanol extracts obtained from *Plantago* species. *J Agric Food Chem* 2005; 53:1927-1933.
23. Felklova M. Antibacterial properties of *Plantago lanceolata* extracts. *Pharm Zentralhalle* 1958; 97:61-65.
24. Chiang LC, Chiang W, Ng LT, Lin CC. Antiviral activity of *Plantago major* extracts and related compounds in vitro. *Antiviral Res* 2002; 55:53-62.
25. Flier H, Verspohl EJ. Antispasmodic activity of an extract from *Plantago lanceolata* and some isolated compounds. *Phytomedicine* 2007; 14:409-415.
26. Nizioł-Łukaszewska Z, Gaweł-Bęben K, Rybczyńska-Tkaczyk K, Jakubczyk A, Karaś M, Bujak T. Biochemical properties, UV-protecting and fibroblast growth-stimulating activity of *Plantago lanceolata* L. extracts. *Industrial Crops and Products* 2019. 138:111453.
27. Parus A, Gryś A. *Plantago lanceolata* L. - healing properties, progress in Phytotherapy. *Postępy fitoterapii* 2010; 3:162–165 (in Polish).
28. Jurisic R, Debeljak Z, Vladimir-Knezevic S, Vukovic J. Determination of aucubin and catalpol in *Plantago* species by micellar electrokinetic chromatography. *Z Naturforsch* 2004; 59c:27-31.
29. Murai M, Tamayama Y, Sansei N. Phenylethanoids in the herb of *Plantago lanceolata* and inhibitory effects on arachidonic acid-induced mouse ear edema. *Planta Med* 1995; 61:479-480.
30. Beara IN, Lesjak MM, Jovin ED, Balog KJ, Anackov GT, Orcić DZ, Mimica-Dukić NM. Plantain (*Plantago* L.) species as novel sources of flavonoid antioxidants. *J Agric Food Chem*. 2009; 57:9268–9273.
31. Demir S, Aliyazicioglu Y, Turan I, Misir S, Mentese A, Yaman SO, Akbulut K, Kilinc K, Deger O. Antiproliferative and proapoptotic activity of Turkish propolis on human lung cancer cell line. *Nutr Cancer* 2016; 68(1):165-72.
32. Tyszka-Czochara M, Paško P, Reczyński W, Szłósarczyk M, Bystrowska B, Opoka W. Zinc and propolis reduces cytotoxicity and proliferation in skin fibroblast cell culture: total polyphenol content and antioxidant capacity of propolis. *Biol Trace Elem Res* 2014; 160(1):123-31.
33. Jacob A, Parolia A, Pau A, Davamani Amalraj F. The effects of Malaysian propolis and Brazilian red propolis on connective tissue fibroblasts in the wound healing process. *BMC Complement Altern Med* 2015; 15:294.
34. Chaitrakoonthong T, Ampornaramveth R, Kamolratanakul P. Rinsing with L-Ascorbic Acid Exhibits Concentration-Dependent Effects on Human Gingival Fibroblast In Vitro Wound Healing Behavior. *Int J Dent*. 2020; 2020:4706418.
35. Deters AM, Schröder KR, Smiatek T, Hensel A. Ispaghula (*Plantago ovata*) seed husk polysaccharides promote proliferation of human epithelial cells (skin keratinocytes and fibroblasts) via enhanced growth factor receptors and energy production *Planta Med* 2005; 71(1):33-9.
36. Kováč I, Ďurkáč J, Hollý M, Jakubčová K, Peržel'ová V, Mučaji P, Švajdlenka E, Sabol F, Legáth J, Belák J, Smetana K Jr, Gál P. *Plantago lanceolata* L. water extract induces transition of fibroblasts into myofibroblasts and increases tensile strength of healing skin wounds. *J Pharm Pharmacol*. 2015; 67(1):117-25.

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**UTICAJ DODATKA VITAMINA C I EKSTRAKTA BOKVICE U EKSTRAKT
PROPOLISA NA VIJABILNOST FIBROBLASTA U ĆELIJSKOJ KULTURI *IN*
*VITRO***

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ABSTRACT

Propolis je pčelinji proizvod izrazito bogat biološki aktivnim supstancama za koje je dokazano da blagotvorno utiču na zdravlje ljudi. Propolis ima široku primenu u tradicionalnoj medicini za lečenje raznih respiratornih ali i kožnih oboljenja. Propolis se može koristiti samostalno kao prečišćeni ekstrakt ili sa dodatkom raznih biljnih ekstrakata i antioksidanasa radi postizanja sinergističkih efekata. Cilj ovog istraživanja je bio da se ispituju efekti različitih ekstrakata propolisa, komercijalno dostupnih na tržištu, na vijabilnost fibroblasta u ćelijskoj kulturi *in vitro*. Ispitali smo efekat tri različita ekstrakta propolisa: čistog ekstrakta propolisa (25%), ekstrakta propolisa (10%) sa dodatkom vitamina C i ekstrakta propolisa (10%) sa bokvicom i dodatkom vitamina C, na vijabilnost L929 fibroblasta, koristeći MTT test. Uočen je koncentracijski-zavisan efekat svih ispitivanih ekstrakata propolisa na vijabilnost fibroblasta. Takođe, uočene su razlike u uticaju ispitivanih ekstrakata na vijabilnost ćelija koje su povezane sa dodatkom ekstraktu propolisa i obrazac je bio drugačiji u nižim u odnosu na više ispitivane koncentracije. Dodatak vitamina C i ekstrakta bokvice ekstraktu propolisa utiču na delovanje čistog propolisa. Upotreba propolisa u kombinaciji sa biljnim ekstraktima i bioaktivnim supstancama može imati blagotvorne efekte, ali je potrebno te kombinacije najpre razmotriti na osnovu indikacija za koje su ovi proizvodi namenjeni i željenih efekata koje je potrebno postići.

Ključne reči: propolis, bokvica, vitamin C, fibroblasti, *in vitro*